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TOPIC HIGHLIGHT

# Biological basis of radiation-induced pulmonary fibrosis

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# **ABSTRACT**

Pulmonary fibrosis is a major radiotherapy-related toxicity, that worsens patients' quality of life, eventually leading to reduced treatment doses and subsequent decreased cancer control probability. Current therapies for radiation-induced pulmonary fibrosis are largely ineffective; thus, intense research has been done to elucidate the pathogenesis of pulmonary fibrosis and develop therapeutic strategies able to mitigate this severe side effect without interfering with anticancer treatment. The development of radiation-induced pulmonary fibrosis involves multifactorial complex processes. The severity of the response to lung irradiation is influenced by the dose of radiation, the volume of the irradiated parenchyma, the simultaneous use of chemotherapy, preexisting lung diseases, and age and genetic predispositions. Moreover, ionizing irradiation activates a cascade of genetic and nongenetic events involving several cell types, such as pneumocytes, endothelial cells, myofibroblasts and immune cells. This complex radiation-induced biological response is mediated by numerous cross-talking signaling pathways and molecules, including the TGF-β system, cytokines, the NF-kB network, and free-radicals. However, despite the recent progress made in understanding the molecular and cellular mechanisms underlying pathophysiology of lung injury after radiation exposure, much remains to be learned about how the various cells and signaling pathways interact. Thus, further studies are needed to identify reliable markers and effective targets, with the aim of developing therapeutic approaches able to prevent or treat this severe condition.

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Key words: Pulmonary fibrosis; Radiotherapy; Myofibroblasts; Inflammatory mediators; Oxidative stress

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# INTRODUCTION

Radiation therapy is an essential part of treatment programs for multiple thoracic malignancies, along with surgery and chemotherapy. The lungs are one of the most radiosensitive organs and the risk of severe radiation-induced side effects, such as lung fibrosis, is often dose-limiting and can significantly compromise the effectiveness of radiotherapy.

The severity of the response to lung irradiation is influenced by several factors. Radiation-induced lung injury is directly related to the total dose of radiation delivered to the lung[1], to the dose per fraction <sup>[2]</sup> and to the radiation dose rate, while inversely correlates with the number of fractions into which the dose is divided<sup>[3]</sup>. Lung injury is also proportionate to the volume of the irradiated parenchyma<sup>[4]</sup>. Usually, the damage increases as the irradiated volume increases and the lung better tolerates a high dose in a small volume than a low dose to the whole lung.

Moreover, the simultaneous use of adjuvant chemotherapy, preexisting lung disease, poor pulmonary function, age and genetic predispositions have been reported as critical factors for the development of pulmonary fibrosis<sup>[5]</sup>.

Usually, the clinical and histological features of radiation-induced lung disease appear weeks after treatment and are divided into early pneumonitis and late fibrosis.

Radiation pneumonitis usually develops after about 1-3 months after radiotherapy and consists of symptomatic changes such as shortness of breath, congestion, cough and fever.

The phase of chronic inflammation and fibrosis occurs more gradually, months to years after radiotherapy<sup>[6,7]</sup>. Pulmonary fibrosis is characterized by fibroblasts and myofibroblasts accumulation, inflammatory cells infiltration, vascular damage and collagen deposition, with consequent scarring and tissue retraction<sup>[6,8]</sup> and may lead to permanent respiratory failure, with impairment of oxygen transfer<sup>[9]</sup>.

Current therapies for preventing or treating radiation-induced pulmonary fibrosis are largely ineffective<sup>[5,10]</sup>, thus elucidating the signaling pathways underlying pulmonary fibrosis development is urgently needed for the identification of new strategies able to treat or mitigate this devastating condition.

# CELLS INVOLVED IN PULMONARY FIBRO-SIS DEVELOPMENT

Immediately following insult due to radiation, a rapid cascade of genetic and nongenetic events is activated<sup>[11-13]</sup>.

It is now well known that fibrosis is not confined to the injured site only, but it is a systemic process involving several cell types, that are recruited to sites of injury<sup>[14]</sup>.

In particular, pneumocytes (both types I and II) and endothelial cells appear to be the cells most susceptible to nongenetic damage <sup>[15]</sup>.

Radiation causes apoptosis and desquamation of epithelial cells from the alveolar walls, and capillary luminal dilatation and congestion, that in turn lead to an increase in vascular permeability and production of interstitial edema, which may evolve in alveolar edema. The attempt of type II pneumocytes to re-establish a functional alveolar epithelium by proliferating and differentiating into the type I cells might even result in an abnormal epithelial repair, typical of lung fibrosis<sup>[16]</sup>. In addition, radiation-induced pulmonary damage is characterized by a large inflammatory cells infiltration and an increase in collagen deposition<sup>[17,18]</sup>, ultimately leading to impairment of gas exchange and lung function<sup>[19]</sup>. The pivotal cellular mediator of pulmonary fibrosis are myofibroblasts, which are the principal producers of collagen, fibronectins and other matrix molecules after injury<sup>[20-22]</sup>.

Myofibroblasts can originate from a variety of sources, including resident interstitial fibroblasts, epithelial and endothelial cells, which adopt fibroblast-like properties through processes of epithelial/endothelial to mesenchymal transition (EMT/EndMT)<sup>[14,23,24]</sup>, and circulating fibrocytes, fibroblast-like cells derived from bone marrow stem cells<sup>[25,26]</sup>.

The infiltrating immune cells recruited following thorax irradiation, principally the alternatively activated macrophages, but also monocytes, lymphocytes, neutrophils and basophils, have been also proposed to play a central role in the pathogenesis of pulmonary fibrosis<sup>[27,28]</sup>, since they secrete cytokines and chemokines that stimulate the differentiation of fibroblasts and other cells into myofibroblasts<sup>[29]</sup>.

Thus, targeting inflammatory cells has been proposed as a therapeutic strategy for radiation-induced lung injuries. For instance,

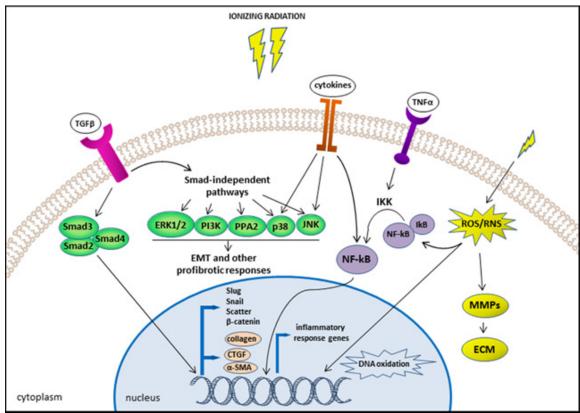


Figure 1 Main signaling pathways involved in the pathogenesis of radiation-induced pulmonary fibrosis.  $\alpha$ -SMA:  $\alpha$  smooth muscle actin; CTGF: connective tissue growth factor; ECM: extracellular matrix; EMT: epithelial to mesenchymal transition; ERK1/2: extracellular-signal-regulated kinases 1/2; IkB: inhibitor of kB; IKK: IkB kinase (responsible of IkB degradation); JNK: c-Jun N-terminal kinases; MMPs: matrix metalloproteinases; NF-kB: nuclear factor-kappa-light-chain-enhancer of activated B cells; p38: p38 mitogen-activated protein kinases; PI3K: phosphatidylinositol 3-kinase; PPA2: protein phosphatase A2; RNS: reactive nitrogen species; ROS: reactive oxygen species; Smad: small mother against decapentaplegic; TGF- $\beta$ : transforming growth factor  $\beta$ ; TNF $\alpha$ : tumor necrosis factor  $\alpha$ 

the beneficial effects of glucocorticoids have been demonstrated in several irradiated organs and tissues<sup>[30]</sup>, and Sivelestat (inhibitor of the neutrophil elastase) has been reported to decrease collagen deposition and neutrophil accumulation in damaged lungs<sup>[31]</sup>.

Dividing cells, such as endothelial and epithelial cells, can also undergo a direct genetic damage, that induces cell apoptosis and a loss of integrity of pulmonary capillaries, leading to alveolar edema and finally to respiratory failure. In addition, it has been reported that various gene loci on chromosomes 1, 17, and 18 influence susceptibility to radiation-induced pulmonary fibrosis<sup>[32,33]</sup>.

# SIGNALING PATHWAYS INVOLVED IN RADIATION-INDUCED PULMONARY FIBROSIS

Ionizing radiation activates a quite complex biological response, mediated by several cross-talking signaling pathways and molecules<sup>[34]</sup> (Figure 1).

#### Cytokines and chemokines

Preclinical models showed that numerous immune cells, such as macrophages, lymphocytes, monocytes, neutrophils and basophils, are recruited into the lung after thorax irradiation [27]. At the same time, an increase in the levels of cytokines and chemokines involved in the recruitment, proliferation and activation of immune cells was observed: macrophage chemoattractant proteins (MCP)-1 and -3 and macrophage inflammatory proteins (MIPs)[35], macrophage-colony stimulating factor (M-CSF), IL-1β[27], interleukin-6 (IL-6)[36], IL-8[37], IL-17[17], CCL11 (Eotaxin)[38] and tumor necrosis factor (TNF)- $\alpha$ [5,39] have all been implicated in lung fibrosis. The use of anti-TNF- $\alpha$  antibodies or TNF- $\alpha$  soluble receptors, for instance, has been suggested as a potential therapeutic strategy to prevent lung fibrosis [40-42].

Among cytokines, Th2-type immune response seem to play a pivotal role in the development of pulmonary fibrosis  $^{[43,44]}$ . The prototypical Th2 cytokine IL-4, whose levels increase during lung fibrosis  $^{[45]}$ , has been shown to exert pro-fibrotic activities, such as inducing the extracellular matrix proteins synthesis, the alternative activation of macrophages and the polarization of T cells towards the Th2 phenotype  $^{[46-48]}$ . Another key profibrotic Th2 cytokine is IL-13  $^{[49]}$ , which promotes the differentiation of fibroblasts into myofibroblasts  $^{[50]}$  and stimulates the transforming growth factor (TGF)- $\beta$  signaling by inducing the production of latent TGF- $\beta$  and enhancing the TGF-activating pathways  $^{[51]}$ .

# TGF-β

The TGF- $\beta$  system is involved in many pathological processes<sup>[52]</sup> and it has been reported to drive the majority of the cellular events associated with radiation-induced fibrosis<sup>[29,53-55]</sup>.

Increases in TGF- $\beta$  serum or plasma levels in patients during and after radiotherapy have been shown to be associated with a higher risk of radiation-induced lung injury<sup>[56-58]</sup>. Increased TGF- $\beta$ 1 and TGF- $\beta$ 2 concentrations have been correlated to the development of fibrosis in a murine model of radiation-induced pulmonary damage <sup>[59]</sup>.

TGF- $\beta$  is produced by mesenchymal, inflammatory (including lymphocytes, macrophages, eosinophils and neutrophils) and epithelial cells<sup>[60-62]</sup>. Moreover, it has been shown that certain bronchial and alveolar cells can be sources of TGF- $\beta$ <sup>[63]</sup>.

TGF- $\beta$  is usually maintained in an inactive state by a latency-associated protein (LAP), but it is rapidly activated by a radiation-induced proteolytic cleavage, that dissociates TGF- $\beta$  from LAP.

Once active, TGF- $\beta$  stimulates the terminal differentiation of progenitor fibroblasts to functional fibrocytes<sup>[21,64]</sup>, induces connective

tissue and extracellular matrix (ECM) deposition<sup>[65]</sup>, the synthesis of matrix-modifying enzymes (i.e. matrix metalloproteinases, MMPs) and stimulates the expression of  $\alpha$ -SMA, a hallmark of myofibroblasts shown to be overexpressed in areas of active fibrosis <sup>[66]</sup>. Moreover, in the immune system, TGF- $\beta$  suppresses T-cell responses<sup>[17]</sup>.

The binding of TGF-β to its specific receptors TGF-βR I and II leads to the induction of the Smad family of transcriptional activators - particularly Smad3<sup>[67]</sup> - that modulate the expression of target genes and induce cellular dedifferentiation and reprogramming by interacting with specific binding sequences in the regulatory regions of the genes<sup>[29,68]</sup>. Smad proteins, in addition, can induce other transcription factors, such as Slug, Snail, Scatter, lymphoid enhancing factor-1, and beta-catenin<sup>[24]</sup>, in turn facilitating EMT.

The participation of TGF- $\beta$ /Smad signaling in the induction of fibrosis was demonstrated by several works, showing that Smad3-deficient animals displayed a reduced fibrotic response<sup>[20,69]</sup> and an accelerated wound healing<sup>[70]</sup> following irradiation and that a hampered Smad activity led to a significant decrease in pulmonary fibrosis development<sup>[71,72]</sup>. Indeed, antibodies against TGF- $\beta$  have been reported to decrease the radiation-induced fibroblasts to fibrocytes differentiation<sup>[73]</sup> and inhibitors of TGF- $\beta$ RI (SM16) or TGF- $\beta$ RI serine/threonine kinase (LY2109761) have shown to reduce radiation-induced pulmonary fibrosis in animal models<sup>[74,75]</sup>.

However, other studies suggested that the therapeutic suppression of the TGF-β/Smad pathway should be limited to the early phases of the disease, since this pathway appears not necessary for the maintenance of the fibrotic phenotype<sup>[76,77]</sup>.

Besides the canonical Smad pathway, TGF- $\beta$  can also activate non-Smad-mediated pathways, such as Rho family proteins, mitogenactivated protein kinases (MAPKs) (ERK1/2, JNK, p38), P13K, protein phosphatase (PP) 2A and the epithelial polarity protein Par6  $^{(78-80)}$ 

It has been reported that the extent and reversibility of EMT was influenced by the cross-talk between the classic TGF- $\beta$  pathway and the above mentioned signaling molecules<sup>[5]</sup>.

Accordingly, a Rho/ROCK inhibitor and statins (modulating the Smad pathway) have shown to decrease radiation-induced lung fibrosis<sup>[81]</sup>.

#### Connective tissue growth factor (CTGF)

CTGF is induced by TGF- $\beta$ , and usually acts as a mediator of TGF- $\beta$  fibrotic effects<sup>[76]</sup>. However, it has been suggested that CTGF, once active, escapes regulation by TGF- $\beta$  and exerts a direct profibrotic effect, contributing to the promotion and maintenance of lung fibrosis <sup>[77,82,83]</sup>. This hypothesis is supported by data demonstrating that CTGF is able to activate collagen type 1 and to exacerbate extracellular matrix synthesis<sup>[84]</sup> and that CTGF deletion in fibroblasts and smooth muscle cells greatly decreases fibrosis<sup>[85]</sup>.

#### NF-kB signaling

The production of cytokines in immune cells is partially regulated at the transcriptional level by DNA binding proteins, such as the transcription factor NF-kB<sup>[86]</sup>, which appears to be a key mediator of the cellular response to irradiation<sup>[87,88]</sup> and to play an important role in the pathogenesis of pulmonary fibrosis<sup>[89,90]</sup>.

NF-kB, usually bound to the inhibitory protein IkB- $\alpha$  in the cytoplasm, is activated by the phosphorylation and degradation of IkB- $\alpha$  induced by several signals, including TNF- $\alpha^{[91,92]}$ , and translocates into the nucleus, where it controls the expression of a number of inflammatory response genes<sup>[93,94]</sup>.

Thus, the suppression of the NF-kB network, either by protecting IkB- $\alpha$  from degradation or by inhibiting the NF-kB-induced upregulation of proinflammatory and profibrogenic cytokines, could be a potential therapeutic approach for preventing or reducing lung fibrosis

#### ECM remodeling

The differentiation of fibroblasts to myofibroblasts during pulmonary fibrosis development lead to an increase in the synthesis of ECM proteins<sup>[95,96]</sup>. However, the changes in ECM composition, in turn, influence myofibroblast differentiation and are determining factors for the progression of the disease.

In addition to the well-known increase in vimentin and alpha-SMA expression induced by irradiation<sup>[97]</sup>, the expression of elastin, type V collagen and tenascin was recently correlated to new collagen formation during lung fibrosis induction<sup>[98]</sup>. Moreover, the observations that MMP-2 and MMP-9 expression increased during radiation-induced lung injury<sup>[99]</sup> and that mice lacking MMP-13 are more resistant to pulmonary fibrosis development<sup>[100]</sup> demonstrated the role of metalloproteinases in the pathogenesis of lung fibrosis.

#### **Surfactant Proteins**

As already mentioned, when the alveolar epithelium is damaged, type II pneumocytes start proliferating to differentiate into type I cells. As a consequence, an increase in the synthesis and release of type II pneumocytes-associated surfactant proteins (SPs) in the circulation is observed<sup>[101]</sup>. In particular, the monitoring of SP-A and SP-D serum levels has been found to be associated with pulmonary fibrosis<sup>[102]</sup> and the SP-C precursor has been suggested as a marker of radiation-induced lung injury<sup>[101]</sup>.

### Oxidative stress

Radiation exposure leads to the production of free-radicals, mostly the reactive oxygen species (ROS, i.e. superoxide, hydrogen peroxide, and hydroxyl radical) and the reactive nitrogen species (RNS). These reactive species cause oxidative stress to the tissue [103,104] and appear to be involved in the molecular pathology of fibrosis [105] in concert with cytokines.

The radiation-induced increase in ROS/RNS levels is, indeed, associated to changes in the cytokine pattern, and leads to lipid peroxidation, DNA and protein oxidation and the activation of fibrogenic signals<sup>[106-108]</sup>.

Since its participation in the pathogenesis of fibrosis, oxidative stress has been explored as a potential therapeutic intervention point. One of the most extensively studied treatment strategies are agents which act as free radical scavengers, such as thiols. In particular, the thiol amifostine has been approved by the FDA for use as a radioprotective agent<sup>[109]</sup> and is clinically used to prevent xerostomia in head and neck cancer patients undergoing radiotherapy<sup>[110]</sup>.

ROS/RNS production can also be targeted by enhancing the activity of antioxidant enzymes, such as superoxide dismutases (SODs) and catalase. Several animal studies reported that the administration of a SOD transgene/liposome delivery vehicle or of SOD-catalase mimetics reduced radiation-induced pulmonary fibrosis by preventing oxidative damage<sup>[103,111,112]</sup>.

#### Renin-angiotensin system

The renin-angiotensin system, whose main function is regulating blood pressure and fluid balance in the body, has been reported to be involved in the pathogenesis of radiation-induced lung injury<sup>[113]</sup>. Thus, this system has been proposed as a potential therapeutic target.

Actually, some angiotensin-converting enzyme (ACE) inhibitors and an angiotensin receptor blocker have demonstrated success in protecting lungs from fibrosis after radiation exposure<sup>[114]</sup>.

#### CONCLUSIONS

Radiation-induced lung fibrosis is a major radiotherapy-related toxicity, that worsens patients' quality of life and can eventually force physicians to limit the dose of cancer treatment.

Thus, intense research has been done to elucidate the molecular pathology of pulmonary fibrosis and develop therapeutic interventions that could prevent or treat this severe condition.

The development of radiation-induced pulmonary fibrosis involves multifactorial complex processes. Despite the recent progress made in understanding the molecular and cellular mechanisms underlying pathophysiology of lung injury after radiation exposure, much remains to be learned about how the various cells and signaling pathways interact.

Thus, further studies are needed to identify reliable markers and effective targets, with the aim of developing therapeutic approaches able to prevent or treat this severe condition.

# **REFERENCES**

- Dörr W, Hendry JH. Consequential late effects in normal tissues. Radiother Oncol 2001; 61: 223-231.
- Mehta V. Radiation pneumonitis and pulmonary fibrosis in nonsmall-cell lung cancer: pulmonary function, prediction, and prevention. *Int J Radiat Oncol Biol Phys* 2005; 63: 5-24.
- Potten CS. The significance of spontaneous and induced apoptosis in the gastrointestinal tract of mice. *Cancer Metas Rev* 1992; 11: 179-195
- Hopewell JW, Trott KR. Volume effects in radiobiology as applied to radiotherapy. Radiother Oncol 2000; 56: 283-288.
- Ding NH, Li JJ, Sun LQ. Molecular Mechanisms and Treatment of Radiation-Induced Lung Fibrosis. *Current Drug Targets* 2013; 14: 1347-1356.
- McDonald S, Rubin P, Phillips TL, Marks LB. Injury to the lung from cancer therapy: Clinical syndromes, measurable endpoints, and potential scoring systems. *Int J Radiat Oncol Biol Phys* 1995; 31: 1187-1203.
- Abid S, Malhotra V, Perry M. Radiation-induced and chemotherapy-induced pulmonary injury. Curr Opin Oncol 2001; 13: 242-248
- Stone HB, Coleman CN, Anscher MS, McBride WH. Effects of radiation on normal tissue: consequences and mechanisms. *Lancet Oncol* 2003: 4: 529-536.
- Sime PJ, O'Reilly KM. Fibrosis of the lung and other tissues: new concepts in pathogenesis and treatment. *Clin Immunol* 2001; 99: 308-319.
- Samah M, El-Aidy Ael-R, Tawfik MK, Ewais MM. Ewais Evaluation of the antifibrotic effect of fenofibrate and rosiglitazone on bleomycin-induced pulmonary fibrosis in rats. *Eur J Pharmacol* 2012; 689: 186-193.
- Rubin P, Finkelstein JN, Shapiro D. Molecular biology mechanisms in the radiation induction of pulmonary injury syndromes: Interrelationship between the alveolar macrophage and the septal fibroblast. *Int J Radiat Oncol Biol Phys* 1992; 24: 93-101.
- Rubin P, Johnston CJ, Williams JP, McDonald S, Finkelstein JN. A
  perpetual cascade of cytokines post irradiation leads to pulmonary
  fibrosis. *Int J Radiat Oncol Biol Phys* 1995; 33: 99-109.
- Vujaskovic Z, Marks LB, Anscher MS. The physical parameters and molecular events associated with radiation-induced lung toxicity. Semin Radiat Oncol 2000; 4: 296-307.
- 14. Andersson-Sjöland A, Nihlberg K, Eriksson L, Bjermer L, Wester-

- gren-Thorsson G. Fibrocytes and the tissue niche in lung repair. *Respir Res* 2011; 12: 76.
- Rubin P, Casarett GW. Clinical Radiation Pathology. 1st ed. *Phila-delphia PA, Saunders* 1968: 1-517.
- Coggle JE. Proliferation of type II pneumonocytes after X-irradiation. Int J Radiat Biol Relat Stud Phys Chem Med 1987; 51: 393-399.
- Wilson MS, Wynn TA. Pulmonary fibrosis: pathogenesis, etiology and regulation. Mucosal Immunol 2009; 2: 103-121.
- 18. Rodemann HP, Bamberg M. Cellular basis of radiation-induced fibrosis. *Radiother Oncol* 1995; 35: 83-90.
- Marks LB, Yu X, Vujaskovic Z, Small W Jr, Folz R, Anscher MS. Radiation-Induced Lung Injury. Seminars in Radiation Oncology 2003; 13: 333-345.
- Wynn TA. Cellular and molecular mechanisms of fibrosis. J Pathol 2008; 214: 199-210.
- 21. Yarnold J, Brotons MC. Pathogenetic mechanisms in radiation fibrosis. *Radiother Oncol* 2010; 97: 149-161.
- Grgic I, Duffield JS, Humphreys BD. The origin of interstitial myofibroblasts in chronic kidney disease. *Pediatr Nephrol* 2012; 27: 183-193.
- Willis BC, du Bois RM, Borok Z. Epithelial origin of myofibroblasts during fibrosis in the lung. Proc Am Thorac Soc 2006; 3: 377-382
- Kalluri R, Neilson EG. Epithelial–mesenchymal transition and its implications for fibrosis. *J Clin Invest* 2003; 112: 1776-1784.
- Quan TE, Cowper SE, Bucala R. The role of circulating fibrocytes in fibrosis. Curr Rheumatol Rep 2006; 8: 145-150.
- Schmidt M, Sun G, Stacey MA, Mori L, Mattoli S. Identification of circulating fibrocytes as precursors of bronchial myofibroblasts in asthma. *J Immunol* 2003; 171: 380-389.
- Cappuccini F, Eldh T, Bruder D, Gereke M, Jastrow H, Schulze-Osthoff K, Fischer U, Köhler D, Stuschke M, Jendrossek V. New insights into the molecular pathology of radiation-induced pneumopathy. *Radiother Oncol* 2011; 101: 86-92.
- Zhang H, Han G, Liu H, Chen J, Ji X, Zhou F, Zhou Y, Xie C. The development of classically and alternatively activated macrophages has different effects on the varied stages of radiation-induced pulmonary injury in mice. *J Radiat Res* 2011; 52: 717-726
- Mancini ML, Sonis ST. Mechanisms of cellular fibrosis associated with cancer regimen-related toxicities. Front Pharmacol 2014; 5:51
- Sekine I, Sumi M, Ito Y, Nokihara H, Yamamoto N, Kunitoh H, Ohe Y, Kodama T, Saijo N, Tamura T. Retrospective analysis of steroid therapy for radiation-induced lung injury in lung cancer patients. *Radiother Oncol* 2006; 80: 93-97.
- Fujino N, Kubo H, Suzuki T, He M, Suzuki T, Yamada M, Takahashi T, Ota C, Yamaya M. Administration of a specific inhibitor of neutrophil elastase attenuates pulmonary fibrosis after acute lung injury in mice. *Exp Lung Res* 2012; 38: 28-36.
- Haston CK, Zhou X, Gumbiner-Russo L, Irani R, Dejournett R, Gu X, Weil M, Amos CI, Travis EL. Universal and radiationspecific loci influence murine susceptibility to radiation-induced pulmonary fibrosis. *Cancer Res* 2002; 62: 3782-3788.
- Haston CK, Travis EL. Murine susceptibility to radiation-induced pulmonary fibrosis is influenced by a genetic factor implicated in susceptibility to bleomycin-induced pulmonary fibrosis. *Cancer Res* 1997; 57: 5286-5291.
- Shrieve DC, Loeffler JS. Human radiation injury. 1st ed. Lippincott Williams & Wilkinson Editor 2011: 1-533.
- Johnston CJ, Williams JP, Okunieff P, Finkelstein JN. Radiationinduced pulmonary fibrosis: examination of chemokine and chemokine receptor families. Radiat Res 2002; 157: 256-265.
- Arpin D, Perol D, Blay JY, Falchero L, Claude L, Vuillermoz-Blas S, Martel-Lafay I, Ginestet C, Alberti L, Nosov D, Etienne-

- Mastroianni B, Cottin V, Perol M, Guerin JC, Cordier JF, Carrie C. Early variations of circulating interleukin-6 and interleukin-10 levels during thoracic radiotherapy are predictive for radiation pneumonitis. *J Clin Oncol* 2005; 23: 8748-8756.
- Car BD, Meloni F, Luisetti M, Semenzato G, Gialdroni-Grassi G, Walz A. Elevated IL-8 and MCP-1 in the bronchoalveolar lavage fluid of patients with idiopathic pulmonary fibrosis and pulmonary sarcoidosis. *Am J Resp Crit Care Med* 1994; 149: 655-659.
- Huaux F, Gharaee-Kermani M, Liu T, Morel V, McGarry B, Ullenbruch M, Kunkel SL, Wang J, Xing Z, Phan SH. Role of Eotaxin-1 (CCL11) and CC chemokine receptor 3 (CCR3) in bleomycin-induced lung injury and fibrosis. *Am J Pathol* 2005; 167: 1485-1496.
- Oikonomou N, Harokopos V, Zalevsky J, Valavanis C, Kotanidou A, Szymkowski DE, Kollias G, Aidinis V. Soluble TNF mediates the transition from pulmonary inflammation to fibrosis. *PLoS One* 2006; 1: e108.
- Wolthuis EK, Vlaar AP, Choi G, Roelofs JJ, Haitsma JJ, van der Poll T, Juffermans NP, Zweers MM, Schultz MJ. Recombinant human soluble tumor necrosis factor-alpha receptor fusion protein partly attenuates ventilator-induced lung injury. *Shock* 2009; 31: 262-266
- Sandalon Z, Bruckheimer EM, Lustig KH, Rogers LC, Peluso RW, Burstein H. Secretion of a TNFR: Fc fusion protein following pulmonary administration of pseudotyped adeno-associated virus vectors. *J Virol* 2004; 78: 12355-12365.
- Przybyszewska M, Miłoszewska J, Rzońca S, Trembacz H, Pyśniak K, Kotlarz A, Swoboda P, Zalewska M, Małecki M. Soluble TNFalpha receptor I encoded on plasmid vector and its application in experimental gene therapy of radiation-induced lung fibrosis. *Arch Immunol Ther Exp (Warsz)* 2011; 59: 315-326.
- Sime PJ. The antifibrogenic potential of PPARgamma ligands in pulmonary fibrosis. J Investig Med 2008; 56: 534-538.
- Brush J, Lipnick SL, Phillips T, Sitko J, McDonald JT, McBride WH. Molecular mechanisms of late normal tissue injury. *Semin Radiat Oncol* 2007; 17: 121-130.
- Buttner C, Skupin A, Reimann T, Rieber EP, Unteregger G, Geyer P, Frank KH. Local production of interleukin-4 during radiationinduced pneumonitis and pulmonary fibrosis in rats: macrophages as a prominent source of interleukin-4. *Am J Respir Cell Mol Biol* 1997; 17: 315-325.
- Sempowski GD, Beckmann MP, Derdak S, Phipps RP. Subsets of murine lung fibroblasts express membrane-bound and soluble IL-4 receptors. Role of IL-4 in enhancing fibroblast proliferation and collagen synthesis. *J Immunol* 1994; 152: 3606-3614.
- Liu T, Jin H, Ullenbruch M, Hu B, Hashimoto N, Moore B, McKenzie A, Lukacs NW, Phan SH. Regulation of found in inflammatory zone 1 expression in bleomycin-induced lung fibrosis: role of IL-4/IL-13 and mediation via STAT-6. *J Immunol* 2004; 173: 3425-3431.
- Lee E, Yook J, Haa K, Chang HW. Induction of Ym1/2 in mouse bone marrow-derived mast cells by IL-4 and identification of Ym1/2 in connective tissue type-like mast cells derived from bone marrow cells cultured with IL-4 and stem cell factor. *Immunol Cell Biol* 2005; 83: 468-474.
- Wynn TA. IL-13 effector functions. Annu Rev Immunol 2003; 21: 425-456.
- Ingram JL, Rice AB, Geisenhoffer K, Madtes DK, Bonner JC. IL-13 and IL-1beta promote lung fibroblast growth through coordinated up-regulation of PDGF-AA and PDGF-Ralpha. FASEB J 2004; 18: 1132-1134.
- Lee CG, Homer RJ, Zhu Z, Lanone S, Wang X, Koteliansky V, Shipley JM, Gotwals P, Noble P, Chen Q, Senior RM, Elias JA. Interleukin-13 induces tissue fibrosis by selectively stimulating and activating transforming growth factor-β1. *J Exp Med* 2001; 194: 809-821.

- Massague J. TGFβ signaling in context. Nat Rev Mol Cell Biol 2012; 13: 616-630.
- Bonniaud P, Margetts PJ, Ask K, Flanders K, Gauldie J, Kolb M. TGF-beta and Smad3 signaling link inflammation to chronic fibrogenesis. *J Immunol* 2005; 175: 5390-5395.
- 54. Anscher MS, Peters WP, Reisenbichler H, Petros WP, Jirtle RL. Transforming growth factorβ as a predictor of liver and lung fibrosis after autologous bone marrow transplantation for advanced breast cancer. *N Engl J Med* 1993; 328: 1592-1598.
- Vujaskovic Z, Groen HJM. Transforming growth factor beta (TGFβ), radiation-induced pulmonary injury and lung cancer. *Int J Radiat Biol* 2000; 76:511-516.
- 56. Anscher MS, Kong F, Andrews K. Plasma transforming growth factorβ1 as a predictor of radiation pneumonitis. Int J Radiat Onc Biol Phys 1998; 41: 1029-1036.
- Kong FM, Ao X, Wang L, Lawrence TS. The use of blood biomarkers to predict radiation lung toxicity: a potential strategy to individualize thoracic radiation therapy. *Cancer Control* 2008; 15: 140-150.
- 58. Fu XL, Huang H, Bentel G, Clough R, Jirtle RL, Kong FM, Marks LB, Anscher MS. Predicting the risk of symptomatic radiation-induced lung injury using both the physical and biologic parameters V(30) and transforming growth factor beta. Int J Radiat Oncol Biol Phys 2001; 50: 899-908.
- Epperly MW, Travis EL, Sikora C, Greenberger JS. Manganese superoxide dismutase (MnSOD) plasmid/liposome pulmonary radioprotective gene therapy: modulation of irradiation-induced mRNA for IL-1, TNF-alpha, and TGF-beta correlates with delay of organizing alveolitis/fibrosis. *Biol Blood Marrow Transplant* 1999; 5: 204-214.
- Assoian RK, Fleurdelys BE, Stevenson HC, Miller PJ, Madtes DK, Raines EW, Ross R, Sporn MB. Expression and secretion of type beta transforming growth factor by activated human macrophages. *Proc Natl Acad Sci* USA 1987; 84: 6020-6024.
- Grotendorst GR, Smale G, Pencev D. Production of transforming growth factor beta by human peripheral blood monocytes and neutrophils. J Cell Physiol 1989; 140: 396-402.
- Thompson HG, Mih JD, Krasieva TB, Tromberg BJ, George SC. Epithelial-derived TGF-beta2 modulates basal and wound-healing subepithelial matrix homeostasis. *Am J Physiol Lung Cell Mol Physiol* 2006; 291: L1277-1285.
- 63. Mattos MD, Kimura ET, Silva MR, Egami MI, Segreto RA, Segreto HR. Latent TGFbeta1 activation in the lung irradiated in vivo. *Rev Assoc Med Bras* 2002; 48: 329-334.
- Herskind C, Rodemann HP. Spontaneous and radiation-induced differentiation of fibroblasts. *Exp Gerontol* 2000; 35: 747-755.
- 65. Strutz F, Zeisberg M, Renziehausen A, Raschke B, Becker V, van Kooten C, Müller G. TGF-beta 1 induces proliferation in human renal fibroblasts via induction of basic fibroblast growth factor (FGF-2). *Kidney Int* 2001; 59: 579-592.
- Zhang HY, Gharaee-Kermani M, Zhang K, Karmiol S, Phan SH. Lung fibroblast alpha-smooth muscle actin expression and contractile phenotype in bleomycin-induced pulmonary fibrosis. Am J Pathol 1996; 148: 527-537.
- Roberts AB, Tian F, Byfield SD, Stuelten C, Ooshima A, Saika S, Flanders KC. Smad3 is key to TGF-beta mediated epithelial-tomesenchymal transition, fibrosis, tumor suppression and metastasis. *Cytokine Growth Factor Rev* 2006; 17: 19-27.
- Derynck R, Zhang YE. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. *Nature* 2003; 425: 577-584.
- 69. Flanders KC, Sullivan CD, Fujii M, Sowers A, Anzano MA, Arabshahi A, Major C, Deng C, Russo A, Mitchell JB, Roberts AB. Mice lacking Smad3 are protected against cutaneous injury induced by ionizing radiation. *Am J Pathol* 2002; 160: 1057-1068.
- 70. Ashcroft G, Yang X, Glick AB, Weinstein M, Letterio JL, Mizel

- DE, Anzano M, Greenwell-Wild T, Wahl SM, Deng C, Roberts AB. Mice lacking Smad3 show accelerated wound healing and an impaired local inflammatory response. *Nat Cell Biol* 1999; 1: 260-266
- Bonniaud P, Kolb M, Galt T, Robertson J, Robbins C, Stampfli M, Lavery C, Margetts PJ, Roberts AB, Gauldie J. Smad3 null mice develop airspace enlargement and are resistant to TGF-beta mediated pulmonary fibrosis. *J Immunol* 2004; 173: 2099-2108.
- Zhao J, Shi W, Wang YL, Chen H, Bringas P Jr, Datto MB, Frederick JP, Wang XF, Warburton D. Smad3 deficiency attenuates bleomycin-induced pulmonary fibrosis in mice. *Am J Physiol* 2002: 282: L585-L593.
- Hakenjos L, Bamberg M, Rodemann HP. TGF-beta1-mediated alterations of rat lung fibroblast differentiation resulting in the radiation-induced fibrotic phenotype. *Int J Radiat Biol* 2000; 76: 503-509.
- Anscher MS, Thrasher B, Zgonjanin L, Rabbani ZN, Corbley MJ, Fu K, Sun L, Lee WC, Ling LE, Vujaskovic Z. Small molecular inhibitor of transforming growth factor-beta protects against development of radiation-induced lung injury. *Int J Radiat Oncol Biol Phys* 2008; 71: 829-837.
- 75. Flechsig P, Dadrich M, Bickelhaupt S, Jenne J, Hauser K, Timke C, Peschke P, Hahn EW, Gröne HJ, Yingling J, Lahn M, Wirkner U, Huber PE. LY2109761 attenuates radiation-induced pulmonary murine fibrosis via reversal of TGFbeta and BMP-associated proinflammatory and proangiogenic signals. *Clin Cancer Res* 2012; 18: 3616-3627.
- Leask A, Holmes A, Black CM, Abraham DJ. Connective tissue growth factor gene regulation. Requirements for its induction by transforming growth factor-beta2 in fibroblasts. *J Biol Chem* 2003; 278: 13008-13015.
- Holmes A, Abraham D, Sa S, Shiwen X, Black CM, Leask A. CTGF and SMADs, maintenance of scleroderma phenotype is independent of SMAD signaling. *J Biol Chem* 2001; 276: 10594-10601.
- Masszi A, Di Ciano C, Sirokmány G, Arthur WT, Rotstein OD, Wang J, McCulloch CA, Rosivall L, Mucsi I, Kapus A. Central role for Rho in TGF-beta1-induced alpha-smooth muscle actin expression during epithelial-mesenchymal transition. *Am J Physiol Renal Physiol* 2003; 284: F911-924.
- Zhang YE. Non-Smad pathways in TGF-β signaling. Cell Res 2009; 19: 128-139.
- Ozdamar B, Bose R, Barrios-Rodiles M, Wang HR, Zhang Y, Wrana JL. Regulation of the polarity protein Par6 by TGFbeta receptors controls epithelial cell plasticity. *Science* 2005; 307: 1603-1609
- Monceau V, Pasinetti N, Schupp C, Pouzoulet F, Opolon P, Vozenin MC. Modulation of the Rho/ROCK pathway in heart and lung after thorax irradiation reveals targets to improve normal tissue toxicity. *Curr Drug Targets* 2010; 11: 1395-1404.
- Sonnylal S, Shi-Wen X, Leoni P, Naff K, Van Pelt CS, Nakamura H, Leask A, Abraham D, Bou-Gharios G, de Crombrugghe B. Selective expression of connective tissue growth factor in fibroblasts in vivo promotes systemic tissue fibrosis. *Arthritis Rheum* 2010; 62: 1523-1532.
- 83. Mori T, Kawara S, Shinozaki M, Hayashi N, Kakinuma T, Igarashi A, Takigawa M, Nakanishi T, Takehara K. Role and interaction of connective tissue growth factor with transforming growth factor-beta in persistent fibrosis: A mouse fibrosis model. *J Cell Physiol* 1999; 181: 153-159.
- Brigstock DR. Connective tissue growth factor (CCN2, CTGF) and organ fibrosis: lessons from transgenic animals. *J Cell Com*mun Signal 2010; 4: 1-4.
- Liu S, Shi-wen X, Abraham DJ, Leask A. CCN2 is required for bleomycin-induced skin fibrosis in mice. *Arthritis Rheum* 2011; 63: 239-246.

- Blackwell TS, Christman WW. The role of nuclear factor-kB in cytokine gene regulation. Am J Respir Cell Mol Biol 1997; 17: 3-9.
- 87. Ahmed KM, Li JJ. NF-kappa B-mediated adaptive resistance to ionizing radiation. *Free Radic Biol Med* 2008; 44: 1-13.
- Hallahan DE, Spriggs DR, Beckett MA, Kufe DW, Weichselbaum RR. Increased tumor necrosis factor alpha mRNA after cellular exposure to ionizing radiation. *Proc Natl Acad Sci USA* 1989; 86: 10104-10107.
- Giri SN, Gurujeyalakshmi G, Wang Y. Suppression of bleomycininduced increased production of nitric oxide and NF-kB activation by treatment with taurine and niacin. Adv Exp Med Biol 2002; 483: 545-561.
- Rabbani ZN, Mi J, Zhang Y, Delong M, Jackson IL, Fleckenstein K, Salahuddin FK, Zhang X, Clary B, Anscher MS, Vujaskovic Z. Hypoxia inducible factor lalpha signaling in fractionated radiation-induced lung injury: role of oxidative stress and tissue hypoxia. *Radiat Res* 2010; 173: 165-174.
- Blonska M, You Y, Geleziunas R, Lin X. Restoration of NFkappaB activation by tumor necrosis factor alpha receptor complextargeted MEKK3 in receptor-interacting protein-deficient cells. *Mol Cell Biol* 2004; 24: 10757-10765.
- Osborn L, Kunkel S, Nabel GJ. Tumor necrosis factor alpha and interleukin 1 stimulate the human immunodeficiency virus enhancer by activation of the nuclear factor kappa B. *Proc Natl Acad Sci USA* 1989; 86: 2336-2340.
- 93. Baeuerle PA, Henkel T. Function and activation of NF-kappa B in the immune system. *Annu Rev Immunol* 1994; 12: 141-179.
- Schreck R, Reiber P, Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NFkB transcription factor and HIV-1. EMBO J 1992; 10: 2247-2258.
- Tomasek JJ, Gabbiani G, Hinz B, Chaponnier C, Brown RA. Myofibroblasts and mechano-regulation of connective tissue remodeling. *Nat Rev Mol Cell Biol* 2002; 3: 349-363.
- Hinz B, Phan SH, Thannickal VJ, Prunotto M, Desmoulière A, Varga J, De Wever O, Mareel M, Gabbiani G. Recent developments in myofibroblast biology: paradigms for connective tissue remodeling. *Am J Pathol* 2012; 180: 1340-1355.
- Nagarajan D, Melo T, Deng Z, Almeida C, Zhao W. ERK/GSK-3beta/Snail signaling mediates radiation-induced alveolar epithelial-to-mesenchymal transition. *Free Radic Biol Med* 2012; 52: 983-992.
- 98. Blaauboer ME, Boeijen FR, Emson CL, Turner SM, Zandieh-Doulabi B, Hanemaaijer R, Smit TH, Stoop R, Everts V. Extracellular matrix proteins: a positive feedback loop in lung fibrosis? *Matrix Biol* 2014; 34: 170-178.
- Yang K, Palm J, König J, Seeland U, Rosenkranz S, Feiden W, Rübe C, Rübe CE. Matrix-Metallo-Proteinases and their tissue inhibitors in radiation-induced lung injury. *Int J Radiat Biol* 2007; 83: 665-676.
- 100. Flechsig P, Hartenstein B, Teurich S, Dadrich M, Hauser K, Ab-

- dollahi A, Gröne HJ, Angel P, Huber PE. Loss of matrix metal-loproteinase-13 attenuates murine radiation-induced pulmonary fibrosis. *Int J Radiat Oncol Biol Phys* 2010; 77: 582-590.
- 101. Almeida C, Nagarajan D, Tian J, Leal SW, Wheeler K, Munley M, Blackstock W, Zhao W. The role of alveolar epithelium in radiation-induced lung injury. PLoS One 2013; 8: e53628.
- Takahashi H, Sano H, Chiba H, Kuroki Y. Pulmonary surfactant proteins A and D: innate immune functions and biomarkers for lung diseases. *Curr Pharm Des* 2006; 12: 589-598.
- Pan J, Su Y, Hou X, He H, Liu S, Wu J, Rao P. Protective effect of recombinant protein SOD-TAT on radiation—induced injury in mice. *Life Sci* 2012; 91: 89-93.
- 104. Tak JK, Park JW. The use of ebselen for radioprotection in cultured cells and mice. Free Radic Biol Med 2009; 46: 1177-1185.
- Kinnula VL, Crapo JD. Superoxide dismutases in the lung and human lung diseases. Am J Respir Crit Care Med 2003; 167: 1600-1619
- Ghafoori P, Marks LB, Vujaskovic Z, Kelsey CR. Radiationinduced lung injury. Assessment, management, and prevention. Oncology (Williston Park) 2008; 22: 37-53.
- Zhao W, Robbins ME. Inflammation and chronic oxidative stress in radiation-induced late normal tissue injury: therapeutic implications. *Curr Med Chem* 2009; 16: 130-143.
- Robbins ME, Zhao W. Chronic oxidative stress and radiationinduced late normal tissue injury: a review. *Int J Radiat Biol* 2004; 80: 251-259.
- Kouvaris J, Kouloulias V, Vlahos L. Amifostine: the first selective-target and broad-spectrum radioprotector. *Oncologist* 2007; 12: 738-747.
- 109. Koukourakis MI, Panteliadou M, Abatzoglou IM, Sismanidou K, Sivridis E, Giatromanolaki A. Postmastectomy hypofractionated and accelerated radiation therapy with (and without) subcutaneous amifostine cytoprotection. *Int J Radiat Oncol Biol Phys* 2013; 85: e7-13.
- 110. Gao F, Fish BL, Szabo A, Doctrow SR, Kma L, Molthen RC, Moulder JE, Jacobs ER, Medhora M. Short-term treatment with a SOD/catalase mimetic, EUK-207, mitigates pneumonitis and fibrosis after single-dose total-body or whole-thoracic irradiation. *Radiat Res Soc* 2012; 178: 468-480.
- 111. Epperly MW, Bray JA, Krager S, Zwacka R, Engelhardt JF, Travis EL, Greenberger JS. Prevention of late effects of irradiation lung damage by MnSOD gene therapy. Gene Ther 1998; 5: 196-208.
- 112. Robbins ME, Diz DI. Pathogenic role of the renin-angiotensin system in modulating radiation-induced late effects. *Int J Radiat Oncol Biol Phys* 2006; 64: 6-12.
- 113. Moltenti A, Moulder JE, Cohen EF, Ward WF, Fish BL, Taylor JM, Wolfe LF, Brizio-Molteni L, Veno P. Control of radiation-induced pneumopathy and lung fibrosis by angiotensin-converting enzyme inhibitors and an angiotensin II type 1 receptor blocker. *Int J Radiat Biol* 2000; 76: 523-532.